Synthesis, Ring Opening, and Glycosidic Bond Cleavage of 3-Methyl-2'-deoxyadenosine

By Tozo Fujii,* Tohru Saito, and Tsuyoshi Nakasaka (Faculty of Pharmaceutical Sciences, Kanazawa University, Takara-machi, Kanazawa 920, Japan)

Summary Methylation of N'-benzyloxy-1-(2-deoxy- β -deoxy- β -deoxy- β -deoxy- β -deoxy- β -deoxy- β -deoxy-

The 3-methyl-2'-deoxyadenosine structure (type 5a) has been assumed to occur in DNA's which had been treated with various methylating agents ¹ Because of the extraordinary instability of its glycosidic bond to acid hydrolysis at the polynucleotide level, ² it is of prime importance to study this part-structure at the nucleoside level. We now record the first synthesis of 3-methyl-2'-deoxyadenosine (5a), which has enlarged the scope of our general method for the synthesis of 3,9-disubstituted adenines, as well as its behaviour toward hydrolysis

In general agreement with previous results, the reaction of 2'-deoxyadenosine 1-oxide with PhCH₂Br in AcNMe₂ and treatment of the benzylated product with NaClO₄ gave the 1-benzyloxy-derivative (1a), mp 1435—1445°C (decomp), in 85% yield. The perchlorate (1a) was converted into the free base by the use of Amberlite IRA-402 (HCO₃⁻) and the base was treated with H₂O at 3—4°C for 8 days to furnish the formamidoimidazole (2a) $\frac{1}{2}$ H₂O (70% yield), mp 138—139°C (decomp) Methylation of (2a) with anhydrous K₂CO₃ and MeI in HCONMe₂ at room

temperature afforded the N-methylformamido-derivative (3a) (69% yield), mp 141—142 °C (decomp), which was hydrogenolysed with Raney Ni and H₂ (1 atm, room temperature, 90 min) in H₂O in the presence of a mol equiv of toluene-p-sulphonic acid (TsOH). The crude (4a) TsOH that formed was treated with a little Et₃N in MeOH at -18 °C for 3 days to produce the desired compound (5a) TsOH [19% yield from (3a)], mp ca 120 °C (decomp), $\lambda_{\rm max}$ (95% EtOH) 272 nm (unstable), $\lambda_{\rm max}$ (H₂O) (pH 1 or 13) unstable, $\lambda_{\rm max}$ (H₂O) (pH 7) 271 nm (ϵ 16900) (unstable), δ [(CD₃)₂SO] 2 28 (3H, s, CMe), 4 19 (3H, s NMe), 8 63 and 8 71 (1H each, s, purine protons), and 9 15 and 9 23 (2H, =NH₂+ or 2 NH)

The (5a) TsOH thus obtained was found to be very When treated with boiling MeOH for 30 min, it gave 3-methyladenine (6)6 in 99% yield. It underwent hydrolysis to (6) much faster in an aqueous acidic solution and rate constants of 0.25 min⁻¹ (half life 2.7 min), $0.039 \, \mathrm{min^{-1}}$ (18 min), and $0.02 \, \mathrm{min^{-1}}$ (35 min) were determined for the hydrolyses at pH 3 34 and 25 °C, pH 5 00 and 37 °C, and pH 700 and 37 °C, respectively contrast the hydrolysis of methylated DNA at 37 °C at pH 50 or 70 was reported to liberate (6) at a rate of $1.0 \times 10^{-3} \, \mathrm{min^{-1}}$ (half life 11.5 h) or $4.8 \times 10^{-4} \, \mathrm{min^{-1}}$ (24 h) 2a We also found that the rate constant for the hydrolysis of the furanosyl-analogue (5b) TsOH3b at pH 3 34 and 25 °C was 6 9 \times 10⁻⁴ min⁻¹ (half life 17 h) Interestingly, the replacement of the ribosyl-group in (5b) TsOH by the 2-deoxyribosyl-group to give

† Satisfactory spectral data and/or elemental analyses were obtained for all the new compounds described

(5a) · TsOH made the glycosidic bond cleavage 360 times faster. In H₂O at pH 8.98 and 25 °C, (5a) · TsOH was slowly converted into (6) in 45 h, during which time the temporary formation of the ring-opened derivative (4a) was observed. Although the ring opening of (5a) · TsOH was similar to that reported3b for (5b) · TsOH, the observed hydrolytic cleavage of the glycosidic bond in alkaline solution was quite notable.

The glycosidic bond of the imidazole-derivative (3a) was also unstable in aqueous acidic solution. On treatment with 0.1 N aq. HCl at room temperature for 3.5 h, (3a) provided (3c) (61% yield) as a glass. The ribosyl-analogue (3b)3b was stable under similar conditions. The structure of (3c) was confirmed by its cyclization with HCl-EtOH to yield (7), m.p. 180-181 °C, identical with a sample synthesized from 3-methyl-6-methylthiopurine⁶ and benzyloxyamine, and by its hydrogenolysis using Raney Ni and H₂ and spontaneous cyclization to give (6) in 84% yield.

We acknowledge support of this work by a Grant-in-Aid for Special Project Research from the Ministry of Education, Science and Culture, Japan.

(Received, 12th May 1980; Com. 509.)

 $a; R^1 = 2\text{-deoxy-}\beta\text{-D-ribofuranosyl}$

b; $R^1 = \beta$ -D-ribofuranosyl **c**; $R^1 = H$

¹ (a) P. D. Lawley and P. Brookes, Biochem. J., 1963, 89, 127; (b) S. Riazuddin and T. Lindahl, Biochemistry, 1978, 17, 2110, and references cited therein.

² (a) G. P. Margison and P. J. O'Connor, Brochim. Brophys. Acta, 1973, 331, 349; (b) A. A. Maxam and W. Gilbert, Proc. Natl. Acad. Sci. U.S.A., 1977, 74, 560.

³ (a) T. Fujii, T. Saito, and M. Kawanishi, Tetrahedron Lett., 1978, 5007; (b) T. Saito and T. Fujii, J. Chem. Soc., Chem. Commun.,

1979, 135.

(a) T. Fujii, C. C. Wu, and T. Itaya, Chem. Pharm. Bull., 1971, 19, 1368; (b) J. A. Montgomery and H. J. Thomas, J. Med. Chem.,

1972, 15, 182; (c) T. Fujii, C. C. Wu, T. Itaya, S. Moro, and T. Saito, Chem. Pharm. Bull., 1973, 21, 1676.

6 (a) H. Klenow and S. Frederiksen, Brochim. Brophys. Acta, 1961, 52, 384; (b) T. Ueda, K. Miura, and T. Kasai, Chem. Pharm. Bull., 1978, 26, 2122.

⁶ J. W. Jones and R. K. Robins, J. Am. Chem. Soc., 1962, 84, 1914.